



Azzawi Dates (*Phoenix dactylifera*) as a Substitute for Corn as an Energy Source in Sheep Diet: *In vitro* Gas Production and Fermentation

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ABSTRACT

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In vitro gas production technique was used in the current study to evaluate Azzawi date (AD) as a substitute for corn grain (CG). The AD was used to replace corn grain at graded levels in the proportion: 0:100 (CG), 25:75 (AD25), 50:50 (AD50), 75:25 (AD75), and 100:0 (AD100). Gas production (GP) was continuously measured by incubating samples in buffered rumen fluid from cannulated sheep and it was recorded at 3, 6, 9, 12, 24, 48 and 72h of incubation. Cumulative GP, kinetics of GP (a, b and c), ammonia nitrogen (NH₃-N) and volatile fatty acids (VFA) concentrations were determined, while metabolizable energy (ME), net energy (NE), organic matter digestibility (OMD), microbial protein (MP) were estimated. The gas volume was increased ($P < 0.05$) at 12h with increasing the substitution levels of AD in the feed, the values ranged between 31.3 and 44.7 ml/200 mg DM. However, it was found to be decreased ($P < 0.05$) at 48 and 72h with increasing substitution levels of AD. There were no differences in the gas production found at 24h. Data of VFA, NE, OMD and MP were similar ($P > 0.05$) among the feeds. The energy (ME; MJ/kg DM) value of AD25 (10.4) and AD50 (10.3) were comparable to that of CG (10.5); however it was reduced ($P < 0.05$) with AD75 and AD100. Data demonstrated that, Azzawi date at the level of 50 g/100g of substrate (*i.e.*, AD50) may have similar energy contents as of corn grain, and it can be used as a source of energy in ruminant diets either alone or in combination with corn grain.

Key words: Azzawi date, Corn grain, Gas production, *In vitro*.

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INTRODUCTION

The price of energy supplements has been increased dramatically in the recent years with the increase of demand of feeds for animals. The increase in feed price encouraged nutritionists to search for cheaper high-energy feed ingredients. In Egypt, production of dates (*Phoenix dactylifera*) leaves behind a sizable amount of by-products in the form of discarded dates and date pits, which are not suitable for packing. Azzawi date which produced in Siwa Oasis can be classified by: poorer quality varieties; discarded date; small shriveled fruit; unripe, seedless; un-pollinated fruits of the date and insect or worms infected date. Farmers in this region use Azzawi date to feed their animals intact or after grinding. Azzawi dates can be used as a source of energy to replace part of the corn grain used in the diets of ruminants as main source of energy.

Al-Dabeeb (2005) found that, diets supplemented with up to 20% dates could be used efficiently in feeding Najdi lambs without adverse effects on growth performance and digestion of nutrients. An earlier report by El-Hag *et al.* (1993), indicated that the inclusion of dates at 15% of the whole DM of ration was associated with an increase in growth rate of Awassi lambs.

The *in vitro* gas production (GP) technique may be a good tool in feed evaluation, to determine the rate and extent of DM degradation, and allows for more control of the experimental conditions than *in vivo* experiments. Fermentation of feeds by rumen micro-organisms produce VFA and Microbial protein (MP) and gases (Blümmel *et al.*, 1997). Estimation of fermentable energy by the *in vitro* GP technique for a range of feedstuffs incubated alone or in combination has been investigated by several authors (Fakhri *et al.*, 1998; Ben Salem *et al.*, 2002). Results of GP obtained by simulated diets in calibrated syringes were parallel and matched the results obtained *in vivo* for digestible OM intake and live weight gain (Blümmel and Ørskov, 1993).

The information available about the inclusion of discarded dates in diets of ruminant in Egypt is very scarce. Therefore, the main objective of the present study was to evaluate the effects of substitution the discarded Azzawi date (AD) for corn grains (CG) as a source of energy in sheep diets using *in vitro* GP technique.

MATERIALS AND METHODS

Feed samples

Samples of corn grain and Azzawi date were collected in triplicate from different areas. Samples of Azzawi date, collected from Siwa Oasis, which lies in the western desert of Egypt 65 km of Libyan borders, were mixed with corn grain (in triplicate) by the following proportion : 0:100 (CG), 25:75 (AD25), 50:50 (AD50), 75:25 (AD75), and 100:0 (AD100). Feed samples were ground in mills to pass a 1 mm sieve prior to chemical analysis, *in vitro* GP measurements and rumen fermentation. The chemical composition of the feeds used for *in vitro* GP is presented in Table 1.

Table 1. Chemical composition (g/kg DM) of corn grain (CG) and Azzawi date (AD)

Parameter	Feedstuff		SEM	P value
	CG	AD		
Ash	15.8 ^b	43.1 ^a	4.53	<0.001
Crude protein	85.3 ^a	74.2 ^b	8.56	0.048
Ether extract	41.6	38.3	5.12	0.082
Nitrogen free extract	831.9 ^a	769.2 ^b	23.60	<0.001
Neutral detergent fibre	114.1 ^b	188.3 ^a	11.63	<0.001
Acid detergent fibre	53.6 ^b	182.8 ^a	10.34	<0.001
Total sugars	82.8 ^b	469.5 ^a	13.49	<0.001

^{a,b}Different superscripts following means in the row indicate differences at P<0.05.

In vitro gas production

In vitro gas production was carried out using the method described by Menke and Steingass (1988). Buffer and mineral solution were prepared and placed in a water bath at 39°C under continuous flushing with CO₂. Both solid and liquid rumen fractions were collected before the morning feeding from three fistulated Barki sheep fed twice daily with diet containing berseem hay (600 g) and concentrate mixture (400 g) ration divided into two equal meals at 8:00 and 16:00 hrs daily. Sheep were supplemented with minerals and had free access to water throughout the experiment. Rumen contents were collected into pre-warmed insulated bottles, pooled among sheep, homogenized in a laboratory blender, filtered through two layers of cheese-cloth and purged with CO₂. The well mixed and CO₂ flushed rumen fluid was added to the buffered rumen fluid solution (1:2 v/v), which was maintained in a water bath at 39°C, and mixed.

Approximately 200 mg DM of finely ground samples were accurately weighed into calibrated glass syringes (100 ml). Buffered rumen fluid (30 ml) was pipetted into each syringe, containing the feed samples, and the syringes were immediately placed into the water bath at 39°C. Three syringes with only buffered rumen fluid were incubated and considered as the blanks. A total of 135 syringes (three syringes of each triplicate sample within each of the five feeds in three runs on different weeks with three syringes as blanks at each run) were incubated for a period of 72h.

The gas production was recorded after 3, 6, 9, 12, 24, 48 and 72h of incubation. Total gas values were corrected for the blank incubation, and reported gas values are expressed in ml per 200 mg of DM. After 72h of incubation, the contents of the incubation syringes were transferred into pre-weighed 50ml centrifuge tubes, rinsed into the tubes and centrifuged. The supernatant was taken for determination of NH₃-N and volatile fatty acids (VFA). Blank 0h samples were also analyzed for NH₃-N and VFA, and these estimates were used to calculate net NH₃-N and net total VFA concentrations.

The gas production fractions were estimated by fitting the mean gas volumes to the exponential equation of Ørskov and McDonald (1979):

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$$\text{Gas (GP)} = a + b [1 - e^{-(c \times t)}],$$

where, GP is the gas production (ml/200 mg DM) at time t , a is the gas production from the immediately soluble fraction, b is the gas production from the insoluble fraction, c is the gas production rate constant for the insoluble fraction (b).

Chemical analysis

Feed samples were analyzed for DM (#934.01), ash (#942.05), N (#954.01), CP (N \times 6.25, #954.01) and ether extract (#920.39) according to AOAC (1997). The neutral detergent fibre (NDF; Van Soest *et al.*, 1991), acid detergent fibre (ADF; #973.18 of AOAC, 1997) analyses used an ANKOM200 Fibre Analyzer unit (ANKOM Technology Corporation, Macedon, NY, USA). The NDF was assayed without use of α -amylase but with sodium sulfite. Both NDF and ADF are expressed without residual ash. The concentration of NH₃-N in rumen fluid was determined by a Kjeldahl distillation method according to AOAC (1997; #954.01), and ruminal total VFA were determined by steam distillation as described by Warner (1964). Total sugars were measured a colorimetric method of Dubois *et al.* (1959).

Calculations and statistical analysis

The energy values were calculated from the amount of gas produced at 24h of incubation with supplementary analysis of CP, ash and ether extract. This approach was developed by the research group in Hohenheim (Germany) and the ME, OMD and NE values in feeds were calculated using equations of Menke *et al.* (1979) and Menke and Steingass (1988).

$$\text{ME (MJ/kg DM)} = 1.06 + 0.157 \times \text{GP} + 0.084 \times \text{CP} + 0.22 \times \text{EE} - 0.081 \times \text{ash}$$

$$\text{OMD (\%)} = 14.88 + 0.889 \times \text{GP} + 0.45 \times \text{CP} + 0.0651 \times \text{CA}$$

$$\text{NE (MJ/kg DM)} = 0.096 \times \text{GP} + 0.0038 \times \text{CP} + 0.000173 \times \text{EE} + 0.54$$

where, GP is 24-h net gas production (ml/200 mg DM), and CP, EE, ash, ME, OMD, NE are crude protein, ether extract, metabolizable energy, organic matter digestibility and net energy, respectively.

Microbial protein was calculated as 19.3g microbial nitrogen per kg OMD according to Czerkawski (1986).

Data of *in vitro* ruminal GP, fermentation parameters, and microbial protein syntheses were analyzed as a complete random design using the GLM option of SAS (2002). Data of each one of the three runs within the same feeds sample were averaged. Mean values of each individual sample within each feed sample (three samples of each) were used as the experimental unit. Turkey's test was used for the multiple comparisons among mean values for the feeds samples used.

RESULTS

The ash, NDF, ADF and total sugars contents were higher ($P < 0.05$) in Azzawi date as compared to corn grain, while the CP and NFE contents were lower ($P < 0.05$) in AD; there was no difference in the fat content between the two feedstuffs (Table 1).

Gas volume was increased ($P < 0.05$) at 12, 48 and 72h with increasing substitution levels of AD in the diets (Table 2). Corn grain (CG) increased ($P < 0.05$) *b* fraction, while it was lowest ($P < 0.001$) with AD100 and their substitutions. In contrast, the intercept *a* fraction (ml/g OM) and rate of fermentation *c* (h^{-1}) were decreased ($P < 0.05$) in CG and increased with increased proportion of AD in the diet.

Table 2. *In vitro* gas production at different time of incubation and gas production kinetics of corn grain, and Azzawi date at different substitution levels

Item	Feeds [†]					SEM	P value
	CG	AD25	AD50	AD75	AD100		
<i>Gas production, ml/200 mg DM</i>							
GP ₃	5.3 ^d	14.3 ^c	15.0 ^c	25.7 ^b	29.3 ^a	0.56	<0.001
GP ₆	15.0 ^d	22.0 ^c	28.0 ^b	38.3 ^a	40.0 ^a	0.53	<0.001
GP ₉	21.0 ^d	29.3 ^c	32.3 ^b	41.0 ^a	42.3 ^a	0.54	<0.001
GP ₁₂	31.3 ^d	34.3 ^c	40.0 ^b	43.0 ^a	44.7 ^a	0.58	<0.001
GP ₂₄	50.3	50.3	50.7	50.0	49.7	0.39	0.485
GP ₄₈	68.3 ^a	64.3 ^b	60.7 ^{bc}	57.0 ^{cd}	54.3 ^d	0.79	<0.001
GP ₇₂	72.0 ^a	68.3 ^{ab}	65.7 ^b	60.0 ^c	57.7 ^c	0.89	<0.001
<i>Gas production kinetics[‡]</i>							
<i>a</i>	1.6 ^c	5.5 ^b	6.5 ^b	20.7 ^a	23.8 ^a	7.36	<0.001
<i>b</i>	69.3 ^a	64.7 ^b	57.8 ^c	37.8 ^d	31.9 ^e	1.01	<0.001
<i>c</i>	0.040 ^e	0.060 ^d	0.067 ^c	0.078 ^b	0.091 ^a	0.0002	<0.001

[†]Contained g/100 g: CG, 100g corn grain; AD25, 25g Azzawi date plus 75g corn grain; AD50, 50g Azzawi date plus 50g corn grain; AD75, 75g Azzawi date plus 25g corn grain; AD, 100g Azzawi date.

[‡]*a*; soluble fraction (ml/g OM), *b*; insoluble fraction (ml/g OM), *c*; production rate (h^{-1}).

^{abcd}Different superscripts following means in the row indicate differences at $P < 0.05$.

Table 3. *In vitro* ruminal fermentation activities, energy contents, organic matter digestibility (OMD) and microbial protein synthesis (MP) of corn grain and Azzawi date at different substitution levels

Item	Feeds [†]					SEM	P value
	CG	AD25	AD50	AD75	AD100		
NH ₃ -N (mg/dL)	16.38 ^a	12.09 ^{ab}	9.57 ^{abc}	7.05 ^b	3.76 ^c	0.818	<0.001
VFA (M.eq/dL)	27.0	26.5	26.1	26.6	26.4	1.06	0.982
ME (MJ/kg DM)	10.5 ^a	10.4 ^{ab}	10.3 ^{ab}	10.1 ^{bc}	9.9 ^c	0.06	0.002
NE (MJ/kg DM)	5.41	5.40	5.44	5.37	5.34	0.039	0.485
OMD (%)	63.55	63.47	63.69	63.03	62.66	0.351	0.276
MP (g/kg DM)	76.7	76.6	76.8	76.0	75.6	0.31	0.073

[†]Contained g/100 g: CG, 100g corn grain; AD25, 25g Azzawi date plus 75g corn grain; AD50, 50g Azzawi date plus 50g corn grain; AD75, 75g Azzawi date plus 25g corn grain; AD, 100g Azzawi date.

^{abc}Different superscripts following means in the row indicate differences at $P < 0.05$.

The ruminal $\text{NH}_3\text{-N}$ concentration was decreased ($P < 0.05$) with increasing the substitution levels of AD with CG. Data on VFA, NE, OMD and MP did not differ among the feeds tested. The energy contents (ME and NE) of CG and AD100 feeds and their substitutions levels, were similar with AD50 although the energy contents were decreased with the other AD levels *versus* GC.

DISCUSSION

Compared to the CG control, inclusion of Azzawi date induced higher gas production during the period from 3 to 12h of incubation. However, during the periods from 48 to 72h, CG showed higher GP than AD100. The differences in the GP are related to the proportion of fermentable substrate of feeds and could be correlated to the observed higher sugars contents in AD (Table 1). Higher fermentation rate and GP in AD containing feeds *vs.* CG was probably due to higher sucrose contents in AD; conversely CG which had a higher starch contents that has lower fermentation rate than the sucrose in the rumen (Sniffen *et al.*, 1992; Adesogan *et al.*, 2005). However, gas is produced mainly when carbohydrates of the feedstuffs are fermented to short chain fatty acids. The rate of GP is associated with the rapid growth phase of microorganisms and in a mixed culture system, the rate of fermentation will be a result of the interactions between the microorganisms present and the manner in which they digest the particular feed within the system (Mauricio *et al.*, 2001).

The total GP of CG and AD100 was found to be 50.3 and 49.7 ml/200 mg DM, respectively, at 24h of incubation. These values are apparently lower than those reported for barley (64-71 ml/200 mg DM), wheat (60-73 ml/200mg DM) and corn grains (60-82 ml/200 mg DM) by Getachew *et al.* (2002) and Umacalilar *et al.* (2002). Gas production reflects all nutrients fermented, soluble as well as insoluble; and fractions that are not fermentable do not contribute to GP. The GP is the result of the fermentation of carbohydrates and the amount of gas produced is affected by the rate of carbohydrate fermentation, the molar proportions of the VFA and amount of VFA produced (Dijkstra *et al.*, 2005). Differences in parameters 'a' and 'c' indicate different fermentation patterns, suggesting that AD is fermented faster, and to a greater extent.

The CP content of CG was higher than AD and our results showed that CG produce higher ($P < 0.05$) in $\text{NH}_3\text{-N}$ *vs.* AD100. This may be explained by the possible difference in the rumen degradability in CP between CG and AD. The increase in $\text{NH}_3\text{-N}$ concentration suggests that CG had high ruminal-degradable protein and thereby the potential to produce higher levels of microbial protein production. Production of $\text{NH}_3\text{-N}$ from *in vitro* GP technique reflects a balance between protein degradation and the uptake of $\text{NH}_3\text{-N}$ for microbial protein synthesis. The principal energy source for the uptake of $\text{NH}_3\text{-N}$ by microorganisms is carbohydrate; therefore the rate of carbohydrate fermentation had a strong relationship with the rate of rumen protein degradation to $\text{NH}_3\text{-N}$ and for microbial protein production (Van Soest, 1982).

Thus it is important that the rate of release of $\text{NH}_3\text{-N}$ matches as closely as possible the release of energy (NRC, 2007). In diets that are rich in protein but low in digestibility, protein degradation rate exceeds carbohydrate fermentation rate and, as energy is the limiting factor for microbial protein synthesis, excess $\text{NH}_3\text{-N}$ is produced (Andrews *et al.*, 1992). The present data of microbial protein was not affected by feeds, being ranged from 75.58 to 76.66 g/kg OMD. This range is higher than that reported in the *in vitro* studies by Van Nevel and Demeyer (1977) which was 18.2-44.6g N/kg OM digested.

The higher GP as well as OMD value of AD feeds could be attributed to relatively low cell wall content as reported by Khan *et al.* (2002). Moreover, the slightly higher GP from fermentation of intact AD feeds compared to CG can be attributed to the presence of higher readily digestible sugars and other short-chain oligosaccharides not present in the extracted fiber. Gas produced is a direct measure of microbial activity and can be a better index of feed ME than an indirect *in vitro* measure based on nutrients fermented. Our values of ME and NE values were lower than that reported by Ismail *et al.* (2005), they calculated ME and net energy for lactation (NE_L) values of corn, wheat, and barley by *in vitro* GP, the values were 13.08, 12.99, and 12.45 MJ/kg DM and 8.30, 8.25, and 7.84 MJ/kg DM, respectively. The ME and NE contents of these feeds were similar to that reported from *in vivo* work of Seker (2002), while NE values were lower than that reported in NRC (2001) tables. Energy values in grains are related to their grain formation, therefore, lower predicted energy value for grains might be attributable to an early stage of harvest (Krishnamoorthy *et al.*, 1995).

CONCLUSION

Supplementation of Azzawi date at the level of 50g per 100g of substrate apparently has similar energy content as that of corn grain and it can be used as a source of energy in concentrate diets for ruminants either alone or in combination with corn grain. However, this readily available cheaper feed should preferably be used in arid tropics for reducing feed cost or for feeding during of the season of nutritional shortage or for covering the maintenance requirements for animals.

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